Metagenomics: An Advanced Approach for Endophyte Study and Prospecting

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ABSTRACT

Endophytes are beneficial microorganisms that reside in different plant tissues. The culturable endophytes are limited so that the knowledge about the microbial communities inside plant tissue is confined. In this study, we concentrated on the metagenomic studies of endophytes that opened a vast knowledge about them. The study of endophytes through the genetic level contributes detailed knowledge about microbial diversity, their composition, functions, and metabolic pathways. This will help to understand their role in plant growth and development. Through metagenomic studies, we can build more information about how endophytes promote plant growth promotion and the different aspects like biofertilization, plant growth hormone production, and stress tolerance. The use of metagenomics gives the importance of plant microbiota to the sustainable use of agriculture and ecological functions.

Keywords: Endophytes, Metagenomics, Biofertilization, Secondary metabolites, Stress tolerance.

INTRODUCTION

Plants are colonised by plenty of microbes in their tissues. Microbial endophytes are microorganisms that reside within plant tissue and complete their life cycle without causing visible harm or disease symptoms.[1] Endophytes encompass a variety of microorganisms, including bacteria, actinomycetes, and fungi.^[2] They are indeed common plant colonizers and play a crucial role in the health and productivity of plants.^[3] Endophytes can interact with plants in a mutualistic, neutral, pathogenic, or beneficial way.^[4,5] The beneficial microbes, called plant growth-promoting endophytes, influence plant survival and functioning, thereby assisting the host plant's overall functioning. This mutualistic relationship exemplifies how plants and microbes work together to optimize survival and performance.^[6,7] According to studies, barely 1% of the endophytes present in nature have

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been cultivated and biotechnologically investigated.^[8-10] Culture-dependent methods have been crucial for the isolation and identification of endophytic bacteria. Only a small percentage of the overall microbial community is culturable, but many other groups are not.^[11] The use of molecular techniques established a tremendous knowledge about the unculturable endophytes and their interactions with the host plant.

The metagenomics approach has indeed revolutionized the study of microbial endophyte communities.^[12] Metagenomics is the genomic analysis of microorganisms through the direct extraction and sequencing of DNA from an assemblage of microorganisms.^[13] The complete taxonomic categorization, as well as the functional and metabolic pathways of the active endophytic microbes, are revealed by metagenomic research.[14] Discovering novel genetic features and processes of plant-microbe interactions is made easier by metagenomics. Metagenomics enables the study of microorganisms easier and more precise regardless of the kind of environmental samples used. Utilizing metagenomics techniques, researchers can analyse the genomes of microorganisms particularly those associated with plants, known as endophytes. It also

enables the identification of potential genes involved in plant growth and physiological processes. This method has aided in the study of many genes that promote plant growth and have either known or unidentified roles.^[15] A potent sequencing tool for microbial research is Next-Generation Sequencing (NGS). This will help in better understanding and describing the microbiome. Next-Generation Sequencing (NGS) technology has been used in the analysis of soil, rhizospheres, and endophytic microbes by providing high-resolution insights into microbial diversity using 16SrRNA amplicon sequencing, sequencing.[16-20] 454 pyrosequencing, and ITS With the aid of this method, it is possible to examine the metagenome of bacterial endophytes and eliminate sequences with plant origin from the entire dataset.^[21] These techniques are important valuable tools to explore entire microbial communities within plant tissues that possess unique properties that contribute to plant growth and health including phytohormone synthesis, cellular metabolism, and nitrogen fixation from plant root internal tissues.^[15] Illumina sequencing, which produces large and error-free data sets, is currently the most used NGS technique.[22]

Better understanding of plant endophytes through metagenomic analysis will lead to develop solutions to environmental problems caused by the use of chemical fertilizers and creating biological control alternatives for treating plant diseases.^[23] Because the plant associated beneficial endophytes can directly affect plant growth and health through the production of phytohormones, enhancing nutrient uptake and phosphate solubilization, and tolerating biotic and abiotic stresses, these bacteria can provide advantages to the host plant. Additionally, they have the capacity to exert indirect action by eliciting a variety of physiological and immunological responses from the host, which may lead to the expansion of a particular microbiome to combat the pathogen infection. This advantageously aroused microbiome can compete for plant resources and niche spaces or restrict the establishment of plant diseases by producing antibiotics, enzymes that break down fungal cell walls, or siderophores, among other things.^[24-26] They generate biologically active metabolites having enormous potential for use in pharmaceutics, medicine, and agriculture.

Metagenomics and Biofertilization

Chemical fertilizers are crucial in ensuring that the world's population's increasing food needs are met. To produce crops with the highest yields possible, the three main commercial fertilizer types, Nitrogen (N), Phosphate (P), and Potassium (K) are employed.^[27]

However, the increasing utilisation of chemical fertilizers in agriculture has a negative effect on ecosystems. Chemical fertilizers are not properly absorbed by plants, causing them to eventually reach water bodies through leaching and causing eutrophication. Additionally, they may affect soils in several ways, diminishing their ability to store water and disrupting the balance of soil fertility.^[28] In addition to being expensive, they produce more greenhouse gases due to fossil fuel use in their production. It has been necessary to create alternative fertilizers for a very long time that are affordable, efficient, and environmentally benign and that function without harming the environment.^[29]

Biofertilization is the process of enhancing plant growth by improving the availability or supply of essential nutrients.^[30] Biofertilizers are typically bacteria or fungi that can fix nitrogen, solubilize phosphorus, potash, and perform other tasks^[31] Numerous Plant Growth-Promoting Bacteria (PGPB) aid in nitrogen fixation and phosphate solubilization. Metagenomic study of microbiomes facilitates more understanding about the genetic basis of these activities and how much they are contributed to plant health and productivity. Several studies proved that the knowledge of endophytes through metagenomic analysis helps for the eco-friendly use of biofertilizers instead of chemical fertilizers and sustainable growth in agriculture.

Metagenomic analysis of sugarcane revealed the presence of uncovering novel *nifH* gene-producing microbes colonizing the different tissues that facilitate the growth through nitrogen fixation.^[32] Their study reports more than 20 of the most abundant N2-fixing microbes and among them some are renowned examples of N2-fixing bacteria like Rhizobium, Azospirillum, Enterobacter, etc. More than 23 bacterial species that contribute plant growth promotion were found in rice from West Bengal's coastal saline zone using 16SrRNA genebased metagenomic analysis. Among them, an isolate of Kosakonia was shown to have both the capacity to fix nitrogen and to promote plant growth properties.^[33] Metagenomic analysis of bacterial endophytes of *Emilia* sonchifolia reported genes nifD, nifK, and nifH for nitrogen fixation.^[34] Metagenomic data of bacterial endophytes of Panax ginseng reported the presence of nitrogen fixation genes. The partial amplification of the nifH gene revealed two bacterial endophytes Stenotrophomonas maltophilia and Agrobacterium tumefaciens, having the nifH gene.^[35]

Both monobasic $(H_2PO_4^{-2})$ and dibasic (HPO_4^{-2}) phosphate are absorbed by plants from the soil. However, 95-99% of the phosphorus in soil is unavailable to plants because it is exist in precipitated,

immobilized, and insoluble forms.^[36] Phosphorus Solubilizing Microorganisms (PSM) are microorganisms that are capable of converting insoluble phosphate molecules into phosphate that is readily available to plants.^[37] This increases plant growth, productivity, and quality. Azotobacter, Bacillus subtilis, Bacillus cereus, Bacillus megaterium, Pseudomonas putida, Pseudomonas aeruginosa, Enterobacter aerogenes, and Micrococcus luteus have all been discovered as P-fertilizers.^[38] Phosphate solubilization activities in an endemic legume Humboldtia brunonis Wall. revealed the presence of Brevibacillus brevis in culture-based studies and metagenomic analysis identified a broader range of phosphate solubilizers including, Bradyrhizobium, Enterobacter, Fusirhizobium, Methylobacterium, Salmonella, Rhizobium, Acidobacterium, etc. but are not responsive to culture-based techniques. ^[39] Role of Plant Growth Promoting Rhizobacteria in suppressive soils against Fusarium oxysporum f. sp. cubense based on metagenomic analysis showed the presence of Pseudomonas, Burkholderia, and Bacillus are known as PGPR that can solubilize phosphate.^[40] The study of rice seedling establishment of rice seedling showed plant growth promotion activities and nine endophytic bacteria showed phosphate solubilization activity.^[41]

Metagenomics and Phytohormone Production

Microorganisms that enhance plant development are known as plant growth-promoting microorganisms, which produce various plant hormones that play a significant role in maintaining plant health and vitality. Plant phytohormones stimulate plant development and serve as growth regulators. Five key phytohormones that serve as Plant Growth Regulators (PGRs) are auxin, gibberellins, cytokinin, ethylene, and abscisic acid. ^[42] Acetobacter, Azotobacter, Herbaspirillum, and Rhizobium have been reported to synthesize auxin, cytokinin, and gibberellins.^[43] Indole-3-Acetic Acid (IAA), is a important plant hormone secreted by endophytes, that supports the growth and development of plants.^[44,45] It plays a crucial role in promoting plant cell elongation, differentiation, and division. It enhances seeds and tubers germination, stimulates roots and xylem formation, improves lateral root initiation, and regulates vegetative growth and adventitious root formation.^[46] Beyond the primary growth functions it also contributes to pigment production and the biosynthesis of metabolites, and regulates plant responses to gravity, light, and fluorescence. It also aids in photosynthetic processes and increases plant tolerance to adverse environmental conditions.[47]

Using the shotgun approach the potential functional genes present in the endophytic microbiomes of maize

roots were studied.^[48] Analysis of the metagenome revealed the presence of genes *ipdC* and *iaaM*, which encode indole-3-pyruvate decarboxylase and tryptophan 2-monooxygenase respectively. IAA production capacity of the endophytic microbiome of *Emilia sonchifolia* is indicated by enzyme coding gene annotations like *ipdC* and *tna*.^[34] Bacterial endophytes isolated from maize through metagenomic analysis have been identified as producers of indole acetic acid including species such as *Sphingobium*, *Burkholderia*, *Chitinophaga*, *Staphylococcus*, and *Bacillus*.^[49]

Metagenomics and Stress Tolerance

Various environmental factors negatively affect plant growth, development, and yield. These include salinity,^[50] temperature,^[51] drought,^[52] the presence of harmful metals or organic contaminants, and other stresses. ^[53] Among these drought has emerged as one of the most significant and increasingly common challenges. It has a profound effect on various physiological and metabolic processes in crop plants and severely affects their development and productivity.^[54] Under drought stress crop plants show slower growth and development due to its profound effects on root physiology, nutrient intake, leaf structure, photosynthetic activity, and seedling germination.^[55,56]

Endophytes help crop plants to adapt environmental challenges, by enhancing the plants to withstand harsh conditions, supporting plant development, suppressing phytopathogens, and activating stress-induced genes in plants that are not typically activated in response to stress.^[57] Studies on Ananas comosus identified several endophytic bacteria including species from the genera Staphylococcus, Bacillus, and Providentia. They can boost drought tolerance, encourage growth, and increase disease resistance.[58] Similar to this, Sandhya et al[59] showed that many endophytic bacterial strains obtained from different crops, helped plants to withstand drought conditions up to a matric potential of 1.02, and also had the ability to promote growth. Inoculating wheat seedlings with the endophytic strain Pantoea alhagi, which was isolated from the plant Alhagi sparsifolia, significantly improved their growth under drought conditions.^[60] The endophyte-treated plants also showed increased accumulation of soluble sugar and reduced levels of malondialdehyde indicating enhanced stress resistance and reduced cellular damage. The endophytic bacterium Bacillus subtilis B26 helped to mitigate drought stress in the grass Brachypodium distachyon by enhancing the expression of stress-induced genes.[61] Additionally, endophytic fungal strains Ampelomyces sp. and Penicillium sp. isolated from soil subjected to drought and high

salinity were found to improve tomato plans tolerance to both salinity and drought.^[62]

Endophytic bacteria help plants combat drought stress by producing volatile compounds, abscisic acid, ACC-deaminase, and IAA.^[63] The endophytic bacterial strain Bacillus amyloliquefaciens has been shown to effectively alleviate salinity stress in plants by significantly increasing the production of abscisic acid after inoculation. Additionally, rice subjected to stressful conditions inoculated with B. amyloliquefaciens also led to notable improvements in the levels of some key antioxidant amino acids, including cysteine, aspartic acid, glutamic acid, phenylalanine, and proline.[64] Similarly, research on the effects of pre-treating maize seed with the endophytic bacteria Corynebacterium hansenii and Bacillus subtilis, demonstrated enhanced growth and physiological responses in maize under drought stress.[65] Pre-treatment with C. hansenii under drought conditions resulted in increased relative water content, leaf proline, and chlorophyll levels. Additionally, under stressful conditions, ACC deaminase (1-aminocyclopropane-1carboxylate) enzymes produced by endophytic microbial strains reduce the amounts of ethylene in plants thereby mitigating stress-related damage.^[66] According to Jaemsaeng^[67] the endophytic bacterial strain *Streptomyces* sp. enhanced rice's ability to withstand salt stress by producing 1-Aminocyclopropane-1-Carboxylate Deaminase (ACCD), which works by converting an ethylene precursor into ammonia and -ketobutyrate, lowering the level of ethylene in plants.

Metagenomic studies aim to delineate the metabolic pathways of genes responsible for both known and novel secondary metabolites allowing bacterial adaptation to harsh salinity.[68] The metagenomic study of two fingermillets cultivars revealed the presence of genes encoding stress response.^[69] Shotgun metagenome analysis of uncultured microbial communities within endophytic bacteria has identified populations of Proteobacteria and Actinobacteria that are potentially involved in plant growth promotion and abiotic stress tolerance.^[70] Additionally, changes in the endophytic bacterial communities of wheat, assessed by 16S rRNA sequencing, have been correlated with variations in drought stress conditions, highlighting the dynamic role of these microbial communities in responding to environmental stress.^[71] The metagenomic studies of the desert plant Senna ilatlica revealed the presence of bacterial endophytes.

Metagenomics and production of secondary metabolites

Endophytes has the ability to produce diverse kinds of bioactive secondary metabolites for their own establishment and also to boost the plant immune system. Production of metabolite is an effective method for biocontrol.^[72] Various metabolites produced by endophytes include hydrolytic enzymes, cyanides, antibiotics, etc. and they are potential sources of different biological activities like antimicrobial, anti-inflammatory, antifungal, anticancerous, etc. Metagenomic techniques play an important role in elucidating the bioactive compounds produced by the endophytes. Unculturable Actinobacteria from the Streptomyces species, which are known to produce a variety of bioactive compounds and antibiotics, have been found in several studies on medicinal plants.^[73,74] The genome of the bacterial endophytes, Streptomyces species, are rich source of bioactive compounds and contains genes that encode the organic substances ecomycins, munumbicins, xiamycins, kakudumycins, and pseudomycins.^[7] The possibility of bacterial endophytes serving as a source of new metabolites has therefore been confirmed.^[75,76] Additionally, in order to defend themselves against diseases, unculturable endophytes also produce enzymes like pectinase.^[77] Like xiamycin from Streptomyces GT2002/1503, the bioactive substances generated by microbial endophytes have antibacterial and antiviral activities. The endophytic fungus Taxomyces andreanae isolated from the bark of Pacific yew produces the novel anticancer and antitumour Paclitaxel (Taxol).^[78]

The gene *hcnB*, which encodes hydrogen cyanide synthase, was identified.^[48] This gene enhances the production of Hydrogen Cyanide (HCN) and HCN has been reported to be useful in the encouragement of the host plant's growth. They also discovered the genes *ubiC* and *ph₂F*, which encode for chorismate lyase and phenazine biosynthesis, respectively. These genes produce the compounds phenazine and 4-hydroxybenzoate, which aid the host's defense mechanisms and biocontrol processes.^[79] Terpenoid is a crucial secondary metabolite involved in defense mechanisms, stress tolerance, and other processes. Metagenome of E. sonchifolia contains genes involved in the mevalonate pathway of terpenoid production.^[34] The use of the metagenomic technique in studying complex microbial communities associated with lignocellulosic energy food crops such as Populus nigra, Eucalyptus camaldeulensis, and Arundo donax has been studied.^[80] They revealed that the plant microorganisms produced cellulase-coding genes, indicating that they could eventually be employed to hydrolyze cellulose and lignocellulose found in the environment. The results of a metagenomic analysis of various watermelon cultivars showed that the microbiome associated with the fruit is involved in the metabolism of carbohydrates and fruit ripening. All cultivars contained numerous genes related to various cellular processes including signal transduction, post-translational modification, carbohydrate metabolism, transcription, intracellular trafficking, and amino acid and energy metabolism. In this study, different families of structurally related glycoside hydrolases, glycosyl transferases, carbohydrate esterases, auxillary activities, and carbohydrate-binding modules of enzymes that degrade, modify, or create glycosidic bonds were identified.^[81]

Metagenomics and Phytoremediation

Adverse environmental conditions are due to the high quantity of pollutants generated through various activities and their removal is economically challenging. Biological agents can be an alternative method to remediate this problem. The breakdown of contaminants using microorganisms is known as phytoremediation or bioremediation. Endophytes contribute important to bioremediation. To remove metals from the soil and encourage plant development and production, endophyte-based bioremediation is applied.^[62] Some bacterial endophytes have the ability to withstand high levels of pollutants while exhibiting Plant growth promotion.^[82] Endophytic bacteria were isolated from Sedum plumbizincicola, a Zn/Cd hyperaccumulator plant. Five of the isolates helped the plant to grow better such as the production of IAA, phosphate and phosphorus solubilization, siderophores, and ACC deaminase activity. Additionally, these bacteria showed high tolerance to metals like Pb, Zn, and Cd.^[83] Bacillus, Pseudomonas, and Arthrobacter are regarded as the three most common genera of bacteria in metal-contaminated environments.^[84] Zn-tolerant rhizobia isolated from Zn mining soil has been reported to promote the growth of Leucaena leucocephala in contaminated environments.^[85] In the study by,^[86] Cu-resistant Kocuria sp. was a potential Plant Growth-Promoting Rhizobacterium (PGPR), which was isolated from the dry tailing of a copper mine.

The bioremediation process of hydrocarboncontaminated soil by microbial communities was revealed by metagenomic analysis through Illumina sequencing.^[87] The structural and functional properties of microbial communities are identified via shotgun metagenomics. Using shotgun metagenomics, identified the 'HM resistome'- a comprehensive collection of all the heavy metal resistance genes- present in agricultural soil with and without cadmium pollution was identified. The study functionally annotated genes involved in the translocation of heavy metals, with P-type ATPases functioning in detoxification and the efflux of cadmium being czcA, czcD, czrA, etc. The analysis also revealed multiple genes associated with resistance to Copper (Cu), Nickel (Ni), Iron (Fe), and Cobalt (Co).^[88] The metagenomic analysis is employed to study the microbial adaptation mechanism in contaminated sediments under natural conditions. The results of the study revealed functional gene annotations that include a significant number of genes related to DNA recombination, DNA damage repair, and heavy-metal resistance. In addition, the study also identified genes involved in the exportation, resistance, and reduction pathways of Mercury (Hg), Lead (Pb), Copper (Cu), Arsenic (As), Zinc (Zn), Cadmium (Cd), and Chromium (Cr).^[89]

Metagenomic analysis was used to study the microbial community and their function involved in cd-contaminated and non-contaminated soil.[90] The study revealed the presence of more metabolic genes in cd-contaminated soil including those encoding for ABC transporters and, genes involved in the detoxification process. Comparative metatranscriptomic and metagenomic analysis has been used to study the Cr (VI) remediation genes of unculturable microbes leading to the identification of six novel genes including gsr and mcr that confer Cr tolerance.^[91] These genes demonstrated high Cr (VI) removal efficiencies and good stability in the treatment of Cr (VI)-contaminated industrial wastewater. Enhanced lead phytoextraction by endophytes from indigenous plants is studied.^[92] The combined use of EDTA and isolated endophytes from indigenous plants was studied. Treatment with microbial inoculum alone and combined use treatment revealed the most effective lead phytoextraction in Brassica juncea and Helianthus annus respectively. They also validated these results through metagenomic analysis and confirmed that plant-microbe interaction is important for phytoremediation.

Metagenomics and Biocontrol

Pests and pathogens destroy plants thereby reducing productivity. This brings out the use of chemicals that contaminate the surroundings. The control of plant diseases and phytopathogens using microorganisms is biocontrol. Bacteria and fungi are used as biocontrol agents. By facilitating physical touch or by using particular pathogen-specific defense mechanisms (hyperparasitism, predation, etc.), they can directly suppress infections. They may also produce lytic enzymes, antibiotics, or other substances that have an indirect effect via enhancing plant defenses. Endophytic microbiota act as biocontrol agents through various mechanisms, such as competition for a niche or substrate, engaging hyperparasitism and predation, synthesizing allelochemicals like antibiotics, lytic enzymes, and siderophores, and including systemic resistance in plants.^[93]

Through metagenomic analysis, microbial populations and their effect on controlling diseases are studied. Comparative metagenomics has been used to uncover the microbial population and biocontrol organisms that act against the plant pathogenic fungus Magnaporthe oryzae.[94] and they identified several microbes with antagonistic potential against M. oryzae, suggesting they could serve as potential biocontrol agents for management of rice blast disease. The microorganisms living in and around banana fields affected by Fusarium wilt disease are studied.^[95] They looked at both plants showing disease symptoms and those without symptoms and investigated the presence of microbiota as biocontrol agents and the production of antagonist compounds against phytopathogens. The results showed the presence of two main potentially beneficial microbes, pseudomondales and streptomycetaceae. Manipulating the population of pseudomonads and streptomyceae in rhizosphere soils could help us to reduce or diminish the disease development in plants, even in the presence of the pathogen. Endophytic strain Bacillus amyloliquefaciens suppresses Fusarium oxysporum infection of Chinese wolfberry by modifying its rhizosphere bacterial community.^[96] The biocontrol potential of B. amyloliquefaciens HSB1 and FZB42 was tested against five fungal pathogens that commonly cause root rot in Chinese wolfberries. Metagenomic sequencing assessed bacterial community shifts in the wolfberry rhizosphere following the introduction of each biocontrol strain. The results indicated that HSB1 and FZB42 differentially altered the abundances of various taxa and positively influenced several functions of the inherent wolfberry rhizosphere bacteria.

CONCLUSION

Scientists are interested in studying endophytes and their interactions with the host plant. The invention of molecular techniques like metagenomics opened a vast area to study microbial organisms. The concept of exploring the whole microbiome of a plant provides various knowledge about them and it also helps to understand the diversity, structure, and functions of these microorganisms. Through metagenomic studies, the contributions of endophytes to the growth and development of plants and the genetic level relatedness can be understood. Metagenomics research can focus on specific genes with known functions. However, recent advancements in Next-Generation Sequencing (NGS) technology, particularly shotgun metagenomics have greatly expanded our ability to study microbial communities. This high-throughput sequencing method allows scientists to analyze the entire genetic makeup of endophytic communities. Nowadays comprehensive views of the genes present in these microbial communities are obtained through metagenomic analysis rather than looking at a few specific genes. Detailed study of endophytes and their functions can lead to future studies in economic and environmentfriendly ways.

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CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

ABBREVIATIONS

NGS: Next-Generation Sequencing; rRNA: Ribosomal Ribo Nucleic Acid; ITS: Internal Transcribed Space; PGPB: Plant growth Promoting Bacteria; PGPM: Plant Growth-Promoting Microorganisms; PGRs: Plant Growth Regulators; IAA: Indole-3acetic acid; ACC: 1-aminocyclopropane-1-carboxylate; HCN: Hydrogen Cyanide; DNA: Deoxy Ribo Nucleic Acid; EDTA: Ethylene Diamine Tetra Acetic acid.

SUMMARY

- Endophytes are organisms that reside within plant tissues in a beneficial way to promote plant growth and productivity.
- Less than one percentage of microbial endophytes will be available for research and study in culture conditions. Metagenomic analysis unveils this hidden microbial world in a broad scenario
- Metagenomic analysis provides more insights into the functional and metabolic pathways, essential for plant-microbe interactive development.
- Multifaceted roles of endophytes in plant growth including the production of plant growth regulators, and assimilation of nutrients will be analyzed in greater detail through metagenomic sequencing

- Metabolic profiling is an advancing area of phytochemical research. Genome-based analysis of endophytes will bring some genes to the limelight which has a greater role in plant metabolic pathways. So, bioactivity studies in cognition of these interactive pathways will be a future perspective in endophyte research.
- Other allied aspects like bioremediation, stress tolerance, and biocontrol will be disclosed in greater detail through metagenome sequencing and analysis.

REFERENCES

- Compant S, Clément C, Sessitsch A. Plant growth-promoting bacteria in the rhizo- and endosphere of plants: their role, colonization, mechanisms involved and prospects for utilization. *Soil Biol Biochem.* 2010;42(5):669-78. https://doi.org/10.1016/j.soilbio. 2009.11.024
- Kaul S, Sharma TK, Dhar M. "Omics" tools for better understanding the plantendophyte interactions. *Front Plant Sci.* 2016;7:955. https://doi.org/10.3389/ fpls. 2016.00955
- Turner TR, James EK, Poole PS. The plant microbiome. Genome Biol. 2013;14(6):1-10. https://doi.org/10.1186/gb-2013-14-6-209
- Adegbeye MJ, Salem AZM, Reddy PRK, Elghandour MMM, Oyebamiji KJ. Waste recycling for the eco-friendly input use efficiency in agriculture and livestock feeding. *Resources Use Efficiency in Agriculture*, 2020;1-45. https:// doi.org/10.1007 /978-981-15-6953-1_1
- Tedersoo L, Bahram M, Zobel M. How mycorrhizal associations drive plant population and community biology. *Science*. 2020;367(6480):1223. https:// doi.org/10.1126/ science.aba1223
- Santoyo G, Moreno-Hagelsieb G, Del Carmen Orozco-Mosqueda M, Glick BR. Plant growth-promoting bacterial endophytes. *Microbiol Res.* 2016;183:92-9. https:// doi.org/10.1016/j.micres.2015.11.008
- Adeleke BS, Ayangbenro A, Babalola OO. Effect of endophytic bacterium, Stenotrophomonas maltophilia JVB5 on sunflowers. Plant Prot Sci. 2022;589(3):185-98. https://doi.org/10.17221/171/2021-PPS
- Abedinzadeh M, Etesami H, Alikhani HA. Characterization of rhizosphere and endophytic bacteria from roots of maize (*Zea mays* L.) plant irrigated with wastewater with biotechnological potential in agriculture. *Biotechnol. Rep.* 2019;21(305). https://doi.org/10.1016/j.btre.2019.e00305
- Huang YH. Comparison of rhizosphere and endophytic microbial communities of Chinese leek through high-throughput 16S rRNA gene illumina sequencing. *J Integr Agric*. 2018;17:359-67. https://doi.org /10.1016/ S2095-3119(17)61731-3
- Zhang C, Tian X, Zhang CS. Diversity and probiotic activities of endophytic bacteria associated with the coastal halophyte *Messerschmidia sibirica*. *Appl Soil Ecol*. 2019;143:35-44. https://doi.org/ 10.1016/j.apsoil.2019.05.030
- Hong CE, Kim JU, Lee JW, Bang KH, Jo IH. Metagenomic analysis of bacterial endophyte community structure and functions in *Panax ginseng* at different ages. *3 Biotech*. 2019;9:1-8. https://doi.org/ 10.1007/s13205-019-1838-x
- Selvin J, Lanong S, Syiem D, De Mandal S, Kayang H, Kumar NS, Kiran GS. Culture-dependent and metagenomic analysis of lesser horseshoe bats' gut microbiome revealing unique bacterial diversity and signatures of potential human pathogens. *Microb Pathog*. 2019;137:103675. https://doi. org/10.1016/j.micpath. 2019. 103675
- Handelsman J. Metagenomics: application of genomics to uncultured microorganisms. *Microbiol. Mol. Biol. Rev.* 2004;68(4):669-85. https://doi. org/10. 1128/mmbr.68.4.669-685.2004
- Forbes JD, Knox NC, Ronholm J, Pagotto F, Reimer A. Metagenomics: the next culture-independent game changer. *Front Microbiol.* 2017;8:1069. https://doi.org/ 10.3389/fmicb.2017.01069
- Adeleke BS, Babalola OO. Biotechnological overview of agriculturally important endophytic fungi. *Horticu Environ Biotechnol.* 2021;62:507-20. https://doi.org/ 10.1007/s13580-021-00334-1

- Campisano A, Antonielli L, Pancher M, Yousaf S, Pindo M, Pertot I. Bacterial endophytic communities in the grapevine depend on pest management. *PLoS One*. 2014;9(11):e112763. https://doi.org/10.1371/journal.pone.0112763
- Akinsanya MA, Goh JK, Lim SP, Ting ASY. Metagenomics study of endophytic bacteria in *Aloe vera* using next-generation technology. *Genom Data*. 2015;6:159-63. https://doi.org/10.1016/j.gdata.2015. 09.004
- Sengupta S, Ganguli S, Singh PK. Metagenome analysis of the root endophytic microbial community of Indian rice (*O. sativa* L.). *Genom Data*. 2017;12:41-3. https://doi.org/10.1016/j.gdata.2017.02.010
- Correa-Galeote D, Bedmar EJ, Arone GJ. Maize endophytic bacterial diversity as affected by soil cultivation history. *Front Microbiol.* 2018;9:484. https://doi.org/ 10.3389/fmicb.2018.00484
- Mashiane AR, Adeleke RA, Bezuidenhout CC, Chirima GJ. Community composition and functions of endophytic bacteria of Bt maize. S Afri J Sci. 2018;114(7-8):88-97. http://dx.doi.org/10.17159/sajs. 2018/20170018
- Fadiji AE, Babalola OO. Metagenomics methods for the study of plantassociated microbial communities: a review. J Microbiol Methods. 2020;170:105860. https:// doi.org/10.1016/j.mimet.20 20.105860
- Gloor GB, Hummelen R, Macklaim JM, Dickson RJ, Fernandes AD, MacPhee R, Reid G. Microbiome profiling by illumia sequencing of combinatorial sequence-tagged PCR products. *PLoS One.* 2010;5:e15406. https://doi. org/10.1371/journal.pone.0015406
- Anguita-Maeso M, Trapero-Casas JL, Olivares-García C, Ruano-Rosa D, Palomo-Ríos E, Jiménez-Díaz RM, Landa BB. Verticillium dahliae inoculation and *In vitro* propagation modify the xylem microbiome and disease reaction to Verticillium wilt in a wild olive genotype. *Front Plant Sci.* 2021;12:632689. https://doi.org/10.3389/ fpls.2021.632689
- Carrión VJ, Perez-Jaramillo J, Cordovez V, Tracanna V, De Hollander M, Ruiz-Buck D, Raaijmakers, JM. Pathogen-induced activation of diseasesuppressive functions in the endophytic root microbiome. *Science*. 2019;366(6465):606-12. https://doi.org/ 10.1126/science.aaw9285
- Song C, Zhu F, Carrión VJ, Cordovez V. Beyond plant microbiome composition: Exploiting microbial functions and plant traits via integrated approaches. *Front Bioeng Biotechnol.* 2020;8:896. https://doi.org/10.3389/ fbioe.2020.00896
- Trivedi P, Leach JE, Tringe SG, Sa T, Singh BK. Plant-microbiome interactions: from community assembly to plant health. *Nat Rev Microbiol.* 2020;18(11):607-21. https://doi.org/10.1038/s41579-020-0412-1
- McGuire S. FAO, IFAD, WFP. The state of food insecurity in the world 2015: Meeting the 2015 international hunger targets: Taking stock of uneven progress. Rome: FAO. 2015. *Adv Nutr.* 2015;6:623-4.
- Ongley ED, Xiaolan Z, Tao Y. Current status of agricultural and rural nonpoint source Pollution assessment in China. *Environ Pollut.* 2010;158:1159-68. https://doi.org/ 10.1016/j.envpol.2009.10. 047
- Mahanty T, Bhattacharjee S, Goswami M, Bhattacharyya P, Das B, Ghosh A, Tribedi P. Biofertilizers: A potential approach for sustainable agriculture development. *Environ Sci Pollut Res.* 2016;24:3315-35. https://doi. org/10.1007/s11356-016-8104-0
- Bashan Y. Inoculants of plant growth-promoting bacteria for use in agriculture. Biotechnol Adv. 1998;16(4):729-70. https://doi.org/10.1016/S0734-9750 (98)00003-2
- Verma M, Mishra J, Arora NK. Plant growth-promoting rhizobacteria: diversity and applications. *Environmental Biotechnology: for Sustainable Future*. 2019;129-73. https://doi.org/10. 1007/978-981-10-7284-06
- Singh RK, Singh P, Sharma A, Guo DJ, Upadhyay SK, Song QQ, Li YR. Unraveling nitrogen fixing potential of endophytic diazotrophs of different *Saccharum* species for sustainable sugarcane growth. *Int J Mol Sci.* 2022;23(11):6242. https://doi.org/10. 3390/ijms23116242
- Kunda P, Dhal PK, Mukherjee A. Endophytic bacterial community of rice (*Oryza sativa* L.) from coastal saline zone of West Bengal: 16S rRNAgene based metagenomics approach. *Meta Gene*. 2018;18:79-86. https://doi. org/10.1016/j.mgene.2018.08.004
- Urumbil SK, Anilkumar M. Metagenomic insights into plant growth promoting genes inherent in bacterial endophytes of *Emilia sonchifolia* (Linn.) DC. *Plant Sci Today*. 2021;8(1):6-16. https://doi.org/10.14719/pst.1357
- Chu LL, Bae H. Bacterial endophytes from ginseng and their biotechnological application. J Ginseng Res. 2022;46(1):1-10. https://doi.org/10.1016/j. jgr.2021.04.004

- Gouda S, Kerry RG, Das G, Paramithiotis S, Shin, HS, Patra JK. Revitalization of plant growth promoting rhizobacteria for sustainable development in agriculture. *Microbiol Res.* 2018;206:131-40. https://doi.org/10.1016/j. micres.2017.08.016
- Joshi RD, Kulkarni SS, Bhate MA, Bhusare DU. Phosphate-solubilizing Endophytic Bacteria Isolation from Maize Plant. Int Res J of Science & Engineering. 2022;10(2):21-6.
- Kumar A, Kumar A, Patel H. "Role of microbes in phosphorus availability and acquisition by plants," *Int J Curr Microbiolo Appl Sci.* 2018;7(5):1344-7. https:// doi.org/10.20546/ijcmas.2018.705.161
- Shendye GV, Thamizhseran N. Endophytic bacterial metagenomics and phosphate solubilization activities in an endemic legume *Humboldtia brunonis* Wall. J Appl Biol Biotechnol. 2022;10(6):51-9. http://doi.org/10.7324/ JABB.2022.100606
- Effendi Y, Pancoro A. Revealing the role of plant growth promoting rhizobacteria in suppressive soils against *Fusarium oxysporum* f. sp. *cubense* based on metagenomic analysis. *Heliyon*. 2021;7(8). https://doi. org/10.1016/j.heliyon.2021.e07636
- Ahumada GD, Gómez-Álvarez EM, Dell'Acqua M, Bertani I, Venturi V, Perata P, Pucciariello C. Bacterial endophytes contribute to rice seedling establishment under submergence. *Front Plant Sci.* 2022;13:908349. https:// doi.org/10.3389/fpls.2022. 908349
- Lopes-Lauri F. Plant Growth regulators. Postharvest Management Approaches for Maintaining Quality of Fresh produce. Springer, Cham. 2016;125-39. https://doi.org/10.1007/978-3-319-23582-0_8
- Verma P, Yadav AN, Khannam KS, Mishra S, Kumar S, Saxena AK. Appraisal of diversity and functional attributes of thermotolerant wheat associated bacteria from the peninsular zone of India. *Saudi J Biol Sci.* 2019;26:1882-95. https://doi.org/10.10 16/j.sjbs.2016.01.042
- 44. Mefteh BF, Chenari Bouket A, Daoud A, Luptakova L, N Alenezi F, Gharsallah N, Belbahri L. Metagenomic insights and genomic analysis of phosphogypsum and its associated plant endophytic microbiomes reveals valuable actors for waste bioremediation. *Microorganisms*. 2019;7(10):382. https://doi.org/10.3390/microorga nisms7100382
- Fadiji AE, Babalola OO. Elucidating mechanisms of endophytes used in plant protection and other bioactivities with multifunctional prospects. *Fron Bioeng Biotechnol.* 2020;8:467. https://doi.org/10. 3389/ fbioe.2020.00467
- Singh R, Dubey AK. Diversity and applications of endophytic actinobacteria of plants in special and other ecological niches. *Front Microbiol.* 2018;9:1767. https://doi.org/ 10.3389/fmicb.2018.01767
- Hassan SE-D. Plant growth-promoting activities for bacterial and fungal endophytes isolated from medicinal plant of *Teucrium polium* L. *J Adv Res.* 2017;8:687-95. https://doi.org/10.1016/j.jare. 2017.09.001
- Fadiji AE, Ayangbenro AS, Babalola OO. Unveiling the putative functional genes present in root-associated endophytic microbiome from maize plant using the shotgun approach. J Appl Genet. 2021;62:339-51. https://doi. org/10.1007/s13353-021-00611 -w
- Fadiji AE, Ayangbenro AS, Babalola OO. Metagenomic profiling of the community structure, diversity, and nutrient pathways of bacterial endophytes in maize plant. *Antonie Van Leeuwenhoek*. 2020;113:1559-71. https://doi. org/10.1007/ s10482-020-01463-w
- Frikha Dammak D, Zarai Z, Najah S, Abdennabi R, Belbahri L, Rateb ME, Maalej S. Antagonistic properties of some halophilic thermoactinomycetes isolated from superficial sediment of a solar saltern and production of cyclic antimicrobial peptides by the novel isolate *Paludifilum halophilum*. *BioMed Res Int*. 2017; https://doi.org/ 10.1155/2017/1205258
- Kumar M, Kumar R, Jain V, Jain S. Differential behavior of the antioxidant system in response to salinity induced oxidative stress in salt-tolerant and salt-sensitive cultivars of *Brassica juncea* L. *Biocatal Agric Biotechnol*. 2018;13:12-9. https://doi.org/10. 1016/j.bcab.2017.11.003
- 52. Forni C, Duca D, Glick BR. Mechanisms of plant response to salt and drought stress and their alteration by rhizobacteria. *Plant Soil.* 2017;410:335-56. https://doi.org/10.1007/s11104-016-3007-x
- Monteiro PHR, da Silva FB, de Abreu CM, da Silva GJ. Plant growth promoting rhizobacteria in amelioration of abiotic stresses: A functional interplay and prospective. *Plant Growth-Promoting Microbes for Sustainable Biotic and Abiotic Stress Management*, Springer, Cham, Switzerland, 2021;25-49. https://doi.org/10.1007/978-3-030-66587-6_2

- Chaves MM, Flexas J, Pinheiro C. Photosynthesis under drought and salt stress: regulation mechanisms from whole plant to cell. *Ann Bot.* 2009;103(4):551-60. https://doi.org/10.1093/aob/mcn125
- Li J, Cang Z, Jiao F, Ba, X, Zhang D, Zhai R. Influence of drought stress on photosynthetic characteristics and protective enzymes of potato at seedling stage. J Saudi Soc Agric Sci. 2017;16:82-8. https:// doi.org/10.1016/j. jssas.2015.03.001
- Zhang Z, Zhang J, Xu G, Zhou L, Li Y. Arbuscular mycorrhizal fungi improve the growth and drought tolerance of *Zenia insignis* seedlings under drought stress. *New Forests*. 2019;50(4):593-604. https:// doi.org/10.1007/s11056-018-9681-1
- Verma H, Kumar D, Kumar V, Kumari M, Singh SK, Sharma VK, Kumar A. The potential application of endophytes in management of stress from drought and salinity in crop plants. *Microorganisms*. 2021;9(8):1729. https:// doi.org/10.3390/microorg anisms9081729
- Jayakumar A, Padmakumar P, Nair IC, Radhakrishnan EK. Drought tolerant bacterial endophytes with potential plant probiotic effects from *Ananas comosus*. *Biologia*. 2020;75:1769-78. https://doi.org/ 10.2478/s11756-020-00483-1
- Sandhya V, Shrivastava M, Ali SZ, Sai Shiva Krishna Prasad V. Endophytes from maize with plant growth promotion and biocontrol activity under drought stress. *Russ Agric Sci.* 2017;43(1):22-34. https://doi.org/10.3103/ S1068367417010165
- Chen C, Xin K, Liu H, Cheng J, Shen X, Wang Y, Zhang L. Pantoea alhagi, a novel endophytic bacterium with ability to improve growth and drought tolerance in wheat. Sci Rep. 2017;7:1-14. https://doi.org/10.1038/srep41564
- Gagné-Bourque F, Mayer BF, Charron JB, Vali H, Bertrand A, Jabaji S. Accelerated growth rate and increased drought stress resilience of the model grass *Brachypodium distachyon* colonized by *Bacillus subtilis* B26. *PLoS One*. 2015;10:e0130456. https:// doi.org/10.1371/journal.pone.0130456
- Morsy M, Cleckler B, Armuelles-Millican H. Fungal endophytes promote tomato growth and enhance drought and salt tolerance. *Plants*. 2020;9(7):877. https:// doi.org/10.3390/plants9070877
- Eid AM, Fouda A, Abdel-Rahman MA, Salem SS, Elsaied A, Oelmüller R, Hassan SED. Harnessing bacterial endophytes for promotion of plant growth and biotechnological applications: an overview. *Plants.* 2021;10(5):935. https://doi.org/ 10.3390/plants10050935
- Shahzad R, Khan AL, Bilal S, Waqas M, Kang SM, Lee IJ. Inoculation of abscisic acid-producing endophytic bacteria enhances salinity stress tolerance in *Oryza sativa*. *Environ Exp Bot*. 2017;136:68-77. https://doi. org/10.1016/j.envexpbot.2017.01.010
- Bodhankar S, Grover M, Reddy G. In planta screening of maize seed endophytic bacteria for potential applications under dryland conditions. *Indian J Dryl Agric Res Dev.* 2019;34(1):53-62. http://doi.org/ 10.5958/2231-6701.2019.0009.5
- Doty SL, Oakley B, Xin G, Kang JW, Singleton G, Khan Z, Vajzovic A, Staley JT. Diazotrophic endophytes of native black cottonwood and willow. Symbiosis. 2009;47:23-33. https://doi.org/10.1007/ BF03179967
- Jaemsaeng R, Jantasuriyarat C, Thamchaipenet A. Molecular interaction of 1-aminocyclopropane-1-carboxylate deaminase (ACCD)-producing endophytic *Streptomyces sp.* GMKU 336 towards salt-stress resistance of *Oryza sativa* L. cv. KDML105. *Sci Rep.* 2018;8:1950. https://doi.org/10.1038/ s41598-018 -19799-9
- Tamosiune I, Baniulis D, Stanys V. Role of endophytic bacteria in stress tolerance of agricultural plants: diversity of microorganisms and molecular mechanisms. *Probiotics in Agroecosystem*. 2017;1-29. https://doi. org/10.1007/978-981-10-4059-71
- Prasannakumar MK, Mahesh HB, Desai RU, Kunduru B, Narayan KS, Teli K, Babu GV Metagenome sequencing of fingermillet-associated microbial consortia provides insights into structural and functional diversity of endophytes. 3 *Biotech*. 2020;10:1-17. https://doi.org/10.1007/s13205-019-2013-0
- Žiarovská J, Medo J, Kysel' M, Zamiešková L, Kačániová M. Endophytic bacterial microbiome diversity in early developmental stage plant tissues of wheat varieties. *Plants*. 2020;9(2):266. https://doi.org/ 10.3390/ plants9020266
- Alsaedi ZS, Ashy RA, Shami AY, Majeed MA, Alswat AM, Baz L, Jalal RS. Metagenomic study of the communities of bacterial endophytes in the desert

plant Senna italica and their role in abiotic stress resistance in the plant. *Braz J Biol.* 2022;82. https://doi.org/10.1590/1519-6984.267584

- Mishra J, Arora NK. Secondary metabolites of fluorescent pseudomonads in biocontrol of phytopathogens for sustainable agriculture. *Appl Soil Ecol.* 2018;125:35-45. https://doi.org/10.1016/ j.apsoil. 2017.12.004
- Purushotham N, Jones E, Monk J, Ridgway H. Community structure of endophytic actinobacteria in a New Zealand native medicinal plant *Pseudowintera colorata* (Horopito) and their influence on plant growth. *Microb Ecol.* 2018;76:729-40. https://doi.org/10.1007/s00248-018-1153-9
- Oberhofer M, Hess J, Leutgeb M, Gössnitzer F, Rattei T, Wawrosch C, Zotchev SB. Exploring actinobacteria associated with rhizosphere and endosphere of the native alpine medicinal plant *Leontopodium nivale* subspecies *alpinum. Front Microbiol.* 2019;2531. https://doi.org/10.3389/ fmicb. 2019. 02531
- Kealey C, Creaven CA, Murphy CD, Brady CB. New approaches to antibiotic discovery. *Biotechnol Lett.* 2017;39:805-17. https://doi.org/10.1007/s10529-017-23 11-8
- Singh M, Kumar A, Singh R, Pandey KD. Endophytic bacteria: a new source of bioactive compounds. 3 *Biotech* 2017;7:1-14. https://doi.org/10.1007/ s13205-017-0942-z
- Singh, R., Pandey, K.D., Singh, M., Singh, S.K., Hashem, A., Al-Arjani, A.B.F., Kumar, A. Isolation and characterization of endophytes bacterial strains of *Momordica charantia* L. and their possible approach in stress management. *Microorganisms*. 2022;10(2):290. https://doi.org/10.3390/ microorga nisms10020290
- Wani, MC., Taylor HL, Wall ME, Coggon P, McPhail AT. Plant antitumor agents. VI. Isolation and structure of taxol, a novel antileukemic and antitumor agent from Taxus brevifolia. J Am Chem Soc. 1971;93(9):2325-7. https://doi. org/10.1021/ja00738a 045
- Enagbonma BJ, Babalola OO. Unveiling plant-beneficial function as seen in bacteria genes from termite mound soil. *J Soil Sci Plant Nutr.* 2020;20:421-30. https://doi.org/ 10.1007/s42729-019-00124-w
- Montella S, Ventorino V, Lombard V, Henrissat B, Pepe O, Faraco V. Discovery of genes coding for carbohydrate-active enzyme by metagenomic analysis of lignocellulosic biomasses. *Sci Rep.* 2017;7(1):42623. https://doi. org/10.1038/srep 42623
- Saminathan T, García M, Ghimire B, Lopez C, (A, Nimmakayala P, Reddy UK. Metagenomic and metatranscriptomic analyses of diverse watermelon cultivars reveal the role of fruit associated microbiome in carbohydrate metabolism and ripening of mature fruits. *Front Plant Sci.* 2018;9:4. https:// doi.org/10.3389/fpls.2018.00004
- Malla MA, Dubey A, Yadav S, Kumar A, Hashem A, Abd_Allah, EF. Understanding and designing the strategies for the microbe-mediated remediation of environmental contaminants using omics approaches. *Front Microbiol.* 2018;9:1132. https://doi.org/10.3389/fmicb.2018.01132
- Dubey A, Malla MA, Kumar A, Dayanandan S, Khan ML. Plants endophytes: unveiling hidden agenda for bioprospecting toward sustainable agriculture. *Cri. Rev Biotechnol.* 2020;40(8):1210-31. https:// doi.org/10.1080/0738855 1.2020.1808584

- Pires C, Franco AR, Pereira SI, Henriques I, Correia A, Magan N, Castro PM. Metal (loid)-contaminated soils as a source of culturable heterotrophic aerobic bacteria for remediation applications. *Geomicrobiol J.* 2017;34(9):760-8. https://doi.org/ 10.1080/01490451.2016.1261968
- Rangel WM, Thijs S, Janssen J, Oliveira Longatti SM, Bonaldi DS, Ribeiro PRA, Moreira FM. Native rhizobia from Zn mining soil promote the growth of *Leucaena leucocephala* on contaminated soil. *Int J Phytoremediation*. 2017;19(2):142-56. https://doi.org/10.1080/15226514.2016.1207600
- Hansda A, Kumar V, Anshumali. Cu-resistant *Kocuria sp.* CRB15: a potential PGPR isolated from the dry tailing of Rakha copper mine. 3 *Biotech*. 2017;7:1-11. https://doi.org/10.1007/s13205-017-0757-y
- Siles JA, Margesin R. Insights into microbial communities mediating the bioremediation of hydrocarbon-contaminated soil from an Alpine former military site. *Appl Microbiol Biotechnol.* 2018;102:4409-21. https://doi. org/10.1007/ s00253-018-8932-6
- Salam LB, Obayori OS, Ilori MO, Amund OO. Effects of cadmium perturbation on the microbial community structure and heavy metal resistome of a tropical agricultural soil. *Bioresour Bioprocess*. 2020;7:1-19. https://doi.org/10.1186/ s40643-020-00314-w
- Chen Y, Jiang Y, Huang H, Mou L, Ru J, Zhao J, Xiao S. Long-term and high-concentration heavy-metal contamination strongly influences the microbiome and functional genes in Yellow River sediments. *Sci Total Environ.* 2018;637:1400-12. https://doi.org/10.1016/j.scitotenv.2018.05.109
- Feng G, Xie T, Wang X, Bai J, Tang L, Zhao H, Zhao Y. Metagenomic analysis of microbial community and function involved in cd-contaminated soil. *BMC microbial* 2018;18:1-13. https://doi.org/10.1186/ s12866-018-1152-5
- Pei Y, Tao C, Ling Z, Yu Z, Ji J, Khan A, Li X. Exploring novel Cr (VI) remediation genes for Cr (VI)- contaminated industrial wastewater treatment by comparative metatranscriptomics and metagenomics. *Sci Total Environ* 742:140435. https://doi. org/10.1016/j.scitotenv.2020.140435
- Pietrini I, Grifoni M, Franchi E, Cardaci A, Pedron F, Barbafieri M, Petruzzelli G, Vocciante M. Enhanced lead phytoextraction by endophytes from indigenous plants. *Soil Syst.* 2021;5:55. https://doi.org/10.3390/soilsystems5030055
- Bolívar-Anillo HJ, Garrido C, Collado IG. Endophytic microorganisms for biocontrol of the phytopathogenic fungus Botrytis cinerea. *Phytochem* Rev. 2020;19:721-40. https://doi.org/10.1007/s11101-019-09603-5
- Prasannakumar MK, Netravathi LM, Mahesh HB, Buela Parivallal P, Puneeth ME, Sathish A, Rohit BS. Comparative metagenomic analysis of rice soil samples revealed the diverse microbial population and biocontrol organisms against plant pathogenic fungus *Magnaporthe oryzae*. 3 *Biotech*. 2021;11(5):245. https://doi.org/10.1007/ s13205-021-02783-y
- Kaushal M, Mahuku G, Swennen R. Metagenomic insights of the root colonizing microbiome associated with symptomatic and non-symptomatic bananas in Fusarium wilt infected fields. *Plants*. 2020;9(2):263. https://doi. org/10.3390/plants9020263
- Uwaremwe C, Yue L, Wang Y, Tian Y, Zhao X, Liu Y, Wang R. An endophytic strain of *Bacillus amyloliquefaciens* suppresses *Fusarium oxysporum* infection of chinese wolfberry by altering its rhizosphere bacterial community. *Front Microbio*. 2022;12:782523. https://doi.org/10.3389/fmicb.20 21.782523

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