

# Assessment of Pathogenic Micro-organisms Associated with Vegetable Salads

Sunil Kumar<sup>1,\*</sup>, Mukesh Yadav<sup>1</sup>, Ashwanti Devi<sup>1</sup>, Mahesh Uniyal<sup>2</sup>, Vikas Kumar<sup>1</sup>, Nirmala Sehrawat<sup>1</sup>, Raj Singh<sup>1</sup>

<sup>1</sup>Department of Biotechnology, Maharishi Markandeshwar (Deemed to be University), Mullana (Ambala), Haryana, INDIA.

<sup>2</sup>Department of MMIT and BM, Maharishi Markandeshwar (Deemed to be University), Mullana (Ambala), Haryana, INDIA.

Submission Date: 22-11-2021; Revision Date: 14-12-2021; Accepted Date: 25-12-2021.

## ABSTRACT

Fruits and salads are the healthy sources of nutrients for many people's regular meals served in restaurants, hotels and in the food points of various educational institutions. However, vegetable salads have been linked with outbreaks in many continents all over the globe, which resulted in several health issues. Reason behind the contamination of salads is primarily the poor hygiene management. Carrot, cucumber, onion, cabbage and tomato samples are most commonly sold salads by street vendors and restaurants and stated as common source of transmission. Aim of this study was to probe the prevalence of most common bacterial strains of the salad samples, which are served in and sold by local street vendors, canteens and restaurants. Present investigation revealed a serious concern of the variable bacterial loads in the salad and vegetable samples. Spreading awareness of the potential health risks were also discussed in the present study, which are associated with poor handling of these salads and vegetables.

**Key words:** Bacterial Culture, Bacteriological Load, Food Contamination, Pathogenic Bacteria, Salad.

## Correspondence:

**Dr. Sunil Kumar,**  
Assistant Profesor,  
Department of  
Biotechnology,  
Maharishi Markandeshwar  
(Deemed to be) University,  
Mullana, Ambala,  
Haryana, INDIA.

Email: sunilhr10h@gmail.com  
ORCID: 0000-0002-0689-2244

## INTRODUCTION

Salad vegetables are eaten raw, without being cooked, and without being washed or peeled, increasing the risk of food poisoning.<sup>[1]</sup> Pathogenic microbes can contaminate vegetables and salads during handling, harvesting, through equipment, and via shipping containers.<sup>[2]</sup> Food poisoning is a big public health problem all over the world.<sup>[3]</sup> Certain high-risk factors are associated with human morbidity, mortality, and economic loss, like; absence of proper cooking, consumption without cleaning or washing the salads.<sup>[4,5]</sup>

In the recent years, requirement for freshly cut and ready to eat (RTE) salads has been improved, owing to the food value and health benefits as well.<sup>[6]</sup> Different

organizations like; WHO suggested and encouraged the usage of salads and vegetables as healthy food.<sup>[7-9]</sup> Health beneficial values has enthralled much attention towards usage of salads.<sup>[7]</sup> Raw vegetable has been a congruent part of salads, which offered a vast variety of fiber inside, minerals, vitamins, and additional phyto-chemicals.<sup>[10]</sup> Salads are a rich source of phyto-nutrients, antioxidants, and have health shielding values to improvize overall human health.<sup>[11,12]</sup> Consumption of fiber rich meal are resulted in lesser risk of coronary cardiac disease, lower concentrations of blood cholesterol, chronic obstructive pulmonary disease (COPD), and in stroke protection, hypertension, abridged blood pressure, control of body weight, regulaton and control of glycemia control, and mentainance of good microflora of gut.<sup>[13,14]</sup> Vegetable salads have been seen with anticancer properties and also prevent cardiovascular disease (CVD) expansion.<sup>[15-17]</sup> Raw vegetables, green vegetables, tomatoes, cruciferous vegetables, carrots, and allium vegetables are different types of vegetables, which have been proven to impart protection against malignancies.<sup>[18]</sup> Phenolics are yet another important element, which is also derived from

## SCAN QR CODE TO VIEW ONLINE



www.ajbls.com

DOI: 10.5530/ajbls.2022.11.1

vegetables in the Mediterranean diet. Flavonoids are the specialized phenolics, which are considered as one of the fundamental bioactive compounds in fetching health promoting effects, like; cancer prevention, pro-apoptotic effects, anti-proliferative action.<sup>[19,20]</sup> Additional benefits like; prevention of rheumatoid arthritis, neurodegenerative diseases (Parkinson's and Alzheimer's disease) are also associated with ample usage of vegetables.<sup>[21-23]</sup> Decline in the risk of chronic disease have also been observed in a few investigations where augmented usage of fruit and vegetable in the diet of older people, which associates a feasible nutritional target with enhanced immune system.<sup>[24]</sup> Routine consumption of vegetable and salads should be given continued attention to improve health and diminish the risk of acquisition of disease.<sup>[25]</sup> Modern guiding principles support an intake of fiber rich diet in doubling and inclusion of plentiful vegetables and fruits servings.<sup>[26,27]</sup> This study was designed to investigate the common bacteriological and parasitological contamination sources and different factors associated with ready to eat and raw vegetables and salads.

#### Food-borne pathogens in vegetables salad:

Coliforms are Gram-negative, facultative anaerobic, oxidase negative, non-spore forming rods, which perform lactose fermentation with gas production after 48 hr at 35–37°C.<sup>[28]</sup> Coliform bacteria can be found in the guts of mammals and humans.<sup>[29,30]</sup> The genera *E. coli*, *Klebsiella*, *Citrobacter*, *Enterobacter*, and some species of *Serratia* together form a composition of entire coliform bacteria.<sup>[31,32]</sup> Many bacteria of nonfecal source are included in the total coliform clad; thus, the faecal coliforms are distinguished from total coliforms just by their ability to thrive at high temperatures of 44.5°C.<sup>[33]</sup> The bacteria total coliforms and *E. coli* are commonly utilized as an indicator of hygiene, particularly in the case of faecal contamination.<sup>[34]</sup> Their existence means that pathogens may be present as a result of human or animal faeces contamination.<sup>[35]</sup> Other coliforms can be found in atmosphere (soil, nutrient-rich waters, rotting plant matter) and faeces as well, and some of them can also thrive in different water delivery systems.<sup>[36]</sup>

Gastroenterological diseases by parasitic contamination of raw vegetables and salads is another vital aspect utilized to monitor the human health.<sup>[37]</sup> Common vegetables are purchased from different markets and street vendors of urban and rural areas. Sediments obtained from vegetables washings are examined by microscopy. Once the culture is grown on artificial media, genomic DNA is isolated from culture samples.

Result from a study showed that 34% of the samples under investigation were found with contamination of one or more parasitic species.<sup>[38]</sup> Typical vegetables harboring parasitic contaminations are lettuce (29.5%), and tarragon leaves (2.3%). In lettuce isolates, *Giardia duodenalis* has been traced most commonly from (23.5%).<sup>[39,37]</sup> Other parasites have also been detected in low frequencies from lettuce salads.<sup>[40]</sup> An urgent need is suggested by multiple investigations for tracing sources of parasitic contamination of salads at high detection level.<sup>[41]</sup>

In 1885, a German paediatrician named Theodor Escherichia discovered *E. coli* in the faeces of a child suffering from diarrhoea. *E. coli* is a gram-negative bacillus, which ferments lactose and glucose and produce steam and acids. In the human colon environment, *E. coli* is the most copious facultative anaerobic flora amongst others that colonises the latter. *E. coli* belongs to the faecal coliform community and is a more precise source of faecal contamination. Not all strains of *E. coli* are commensals, a few strains have shown the pathogenic properties, which become lethal in human, avian and other mammalian diseases and also in food-borne complications.<sup>[42]</sup> Some *E. coli* strains develop virulence traits that enable them to cause a wide range of diseases. There are many types of pathogenic *E. coli* strains that cause diarrheal outbreaks.<sup>[43]</sup> Differences in three structural antigens are used to classify different strains of *E. coli*: antigens H, O, and K. Human *E. coli* can be divided into three classes based on genetic and clinical criteria: pathogenic (enteric or diarrheagenic), commensal, and extraintestinal pathogenic *E. coli* (ExPEC).<sup>[44,45]</sup>

Diarrhoea causing most frequent agents in infants is *E. coli* (Diarrheagenic) strains, which are categorized under the EPEC (entero-pathogenic *E. coli*), ETEC (entero-toxigenic *E. coli*), a leading cause of travellers' diarrhoea, and enteropathogenic *E. coli* (EPEC, a major cause of travellers' diarrhoea.<sup>[46,47]</sup> Enterohaemorrhagic *E. coli* (EHEC), causes hemolytic uremic syndrome and hemorrhagic colitis, enteroinvasive *E. coli* (EIEC), causes severe dysentery.<sup>[48,49]</sup> DAEC (diffusely adherent *E. coli*) and EAEC (entero-aggregative *E. coli*), are other toxin-producing types of *E. coli*.<sup>[50]</sup>

#### Microbiological investigation

##### Total Plate Count Method

SPC (Standard Plate Count) is one of the most frequently used tests to assess the quality of food in terms of microbial load. This is also recognized as 'total viable count' or 'aerobic plate count'.<sup>[51]</sup> However, the importance of SPCs, varies significantly depends up

on food item types and the its undergoing processing. In SPC methods, vegetable salads dilutions of  $10^{-2}$ ,  $10^{-3}$ , and  $10^{-4}$  are prepared using searial dilution with the help of sterile pipettes. Each dilution is pipetted into sterile Petri dishes in duplicate, with correct labeling of different concentrations. The dilution bottle is stirred atleast 5 times and allowed to rest for at least 3 min before being pipette into the Petri dish. 0.1 ml of each of dilution is transferred aseptically onto the nutrient/selective agar plate surface. A sterile L-shaped glass rod is used to spread the inoculum on the agar plate. Then at 35°C, the plates are incubated for 24-48 hr. Next day, total number of isolated colonies are counted and the total aerobic microorganism per gram are determined. Different dilution factors ( $10^{-2}$ ,  $10^{-3}$  and  $10^{-4}$ ) of vegetable salads samples were utilized for claculation of clony forming units (CFUs/ml). Each of the experiment are performed in duplicate from dilutions to colony count to rule out the handling error. Microbiological qualities of RTE salads and foods are assessed using microbiological guidelines to assist in the analysis and interpretation.<sup>[52]</sup>

### Microbiological Quality Categories

Based on SPC, indicator organism levels, presence or number of pathogenic microbes, three microbiological rating divisions have been allocated (Figure 1). There are four levels of satisfaction: potentially dangerous, unsatisfactory, marginal, and optimal.<sup>[53]</sup>

- Potentially Dangerous – quantities in this range have the potential to cause food poisoning, and prompt action should be taken.
- Unsatisfactory – In this level, results are indicative of poor handling practices or hygiene and hence considered unacceptable food under microbiological limits.
- Marginal – Food items in this level are on the borderline of permissible range of microbiological quality limits, however, issues of hygiene cannot be ruled out.
- Satisfactory – This level states a satisfactory rank of microbiological quality.

### Risk associated with contaminated vegetables salads:

Gastroenteritis in several cases of human have been attributed due to ingestion of contaminated vegetables in the last decade, with a rise in frequency.<sup>[54]</sup> Risk of contracting food-borne illness have mounted as consequence cosumption of raw or improper cooked vegetables, and hence considered as well recognised

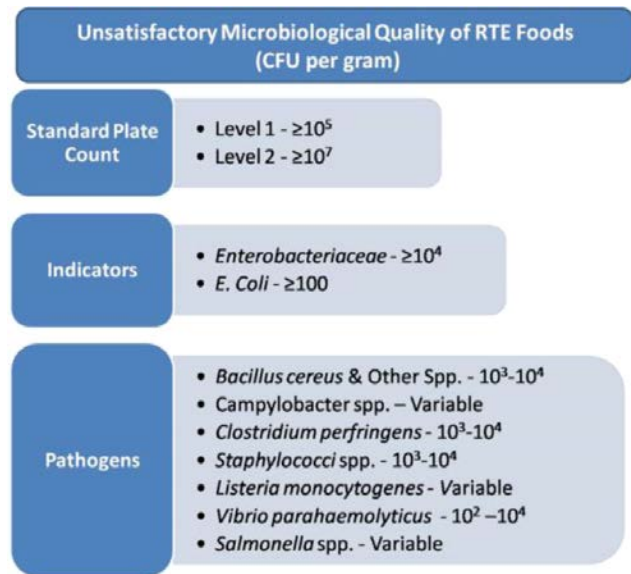
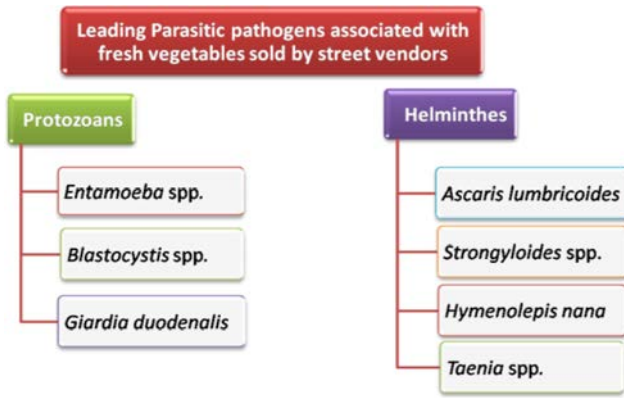


Figure 1: CFUs of different indicator and Pathogenic bacteria showing unsatisfactory microbiological quality of ready-to-eat Foods.

medium for the spread of pathogenic enteric bacteria in human.<sup>[47]</sup>

According to a recent study, vegetable salads like coriander leaf, cucumber, carrots, and tomato are often tainted with *Bacillus cereus*, *Aeromonas* spp., *Campylobacter*, *Salmonella* spp., *Escherichia coli*, *Staphylococcus* spp and *Shigella* spp.<sup>[55]</sup> An escalating graph of food-borne sickness has been seen due to under-cooked or raw vegetables resulting in the tansmission of most of the human gastrointestinal pathogens.<sup>[56]</sup> For safe management of food and to decrease the food associated diseases, various tools have been developed for risk analysis.<sup>[57]</sup> Both the food processor industries and regulatory authorities utilize such tools to control the microbial risk to ensure microbiological quality control and safety for consumers.<sup>[58]</sup> Among various foremost elements of risk analysis, both statistical and scientific information are employed for risk assessment and estimation of probability of death and severe illness.<sup>[58,59]</sup> QMRA (Quantitative microbial risk assessment) is very crucial for the detection of microbial risks associated with specific food consumption, which also helps in providing the information of infection level coupled with a specific food intake.<sup>[60]</sup> This makes sure that the resources are decisively synchronized to curb the risk associated with food-borne microbes.

While the spoiling yeasts, bacteria, and moulds prevail in the raw vegetables microflora, other pathogenic parasites such as helminths, and protozoa with a capability of facilitating infections in human have also been recorded



**Figure 2: Leading Parasitic Pathogens of RTE Vegetable Salad.**

(Figure 2). Minimally processed vegetable salads such as raw vegetables, infected by pathogenic bacteria, like; such as *Listeria monocytogenes*, *Salmonella*, and *E. coli*, may serve as carriers of bacterial agents to humans and they cause food safety issues, especially gastroenteritis as well as being vectors for traveler's diarrhoea and enterotoxigenic.<sup>[27]</sup>

## Identification

### Gram Staining

Following incubation, individual bacterial colonies on each plate are examined and registered based on their form, colour, texture, border, and phenotypic characteristic appearance. An isolated representative colony of the cultured sample is Gram stained. Gram staining is used to expose the cells' characteristic grouping and organisation. The gram stain is also known as differential stain as it has got the discriminatory power to distinguish and stain both Gram negative and Gram positive simultaneously on a single slide.<sup>[61]</sup> Purple color is acquired by Gram positive bacteria (GPB) due to its higher affinity towards crystal violet. When iodine is added, they maintain the crystal violet and purple color is retained after decolorization with acetone. However, Gram negative bacteria (GNB) lose crystal violet color when decolorized with acetone and stain pink due to counterstain Safranin.

### Molecular Methods (MALDI-TOF MS)

MALDI-TOF MS (matrix-assisted laser desorption ionization-time of flight mass spectrometry) has recently emerged as a powerful method for routine clinical isolate detection.<sup>[62]</sup> Bacterial detection using MALDI-TOF MS has been tested quicker, less expensive more precised than traditional phenotypic or molecular approaches.<sup>[63]</sup> The manufacturer's adjusted score values is used to classify MALDI-TOF

MS identifications: a score value of  $\geq 2$  suggested correct species recognition, a score value from 1.7 to 1.9 suggested the identification up to genus level, and a value of 1.7 indicated no identification or not reliable results. In this technique, overnight grown culture are picked from media plate and a thin smear of the colony is prepared over a steel plate of MALDI-TOF MS instrument. The sample is air dried for a while and  $1\mu\text{l}$  of matrix (alpha-cyano-4-hydroxycinnamic aci is added over the pre-dried smear followed by air dry. Now plate with prepared smear is inserted inside the instrument and instrument identified the microorganism using laser beams and analyzing softwares.<sup>[64]</sup>

## DISCUSSION

The present study depicted the microbial variety and their burden in salad vegetables items sold in different canteens and restaurants and isolate and identify bacteria from salad vegetables items. High total viable count (TVC) indicates unsafe condition and therefore the occurrence of possible contamination. Data on vegetable spoiling bacteria and their survival potential is quite obscure and such informations could be of critical importance to determine vegetable shelf life and storage period, and enhance food quality and safety. The current investigation targetted the capacity of vegetable salad to either harbor or inhibit the progression of vegetable spoiling microbes. It is also critical to verify the ability of such microbes to cause pathogenicity to host due to consumption of these spoiled salads and vegetables to avoid the risk of food poisoning. As a result, the period of alive and cultivable microbes in vegetable salad samples are measured with suggestive value of examining product consistency and its ability to sustain micro-organisms. Therefore, present investigation attempted to provide the information on causative agents of food borne illnesses due to contaminated vegetable salads intake.

Finally, showing the potential of vegetables to affect microbial growth would go a long way toward ensuring food consistency, stability, and shelf life. A number of surveillance programs have been conducted previously to draw the demographic scenario of microbial contamination of salad vegetables. Despite of that, informations of contaminants' survival is still limited. Multiple antibiotic resistances were found among bacterial isolates on salad vegetables, according to previous report. Thus, intensive surveillance of isolates to detect emerging developing world is needed. Since salads do not need any additional heat treatment, vigorously washing of vegetables salads and

treatment of the latter with antibacterial chemicals for a comprehensive period of time is necessary to remove pathogens and dramatically reduce the microbial load. Since the way vegetables were handled on wet markets was less hygienic, it was obvious that samples from wet markets yielded a higher proportion of bacteria. The surroundings and locations for vegetable displays in markets were filthy, and the handlers did not wear gloves when handling the vegetables. Contamination may occur as a result of improper handling and transporting in a contaminated container. Aside from that, market vegetables may have a long holding period, which can lead to the accumulation of pathogenic bacteria.

## CONCLUSION

Finally, we conclude with the following recommendations for vegetables salad: Good hygiene practices should be promoted among all food services providers. Implementing and monitoring of safety regulators should be on top priority. Routine review of vegetables agricultural practices must be followed. Setting up local guidelines and food standards on local data are encouraged. Further studies are recommended on Salads and such food to trace sources of contamination. Association of transfer of antibiotic resistance with food borne bacteria are highly recommended to avoid the spread of the latter in food chain.

## ACKNOWLEDGEMENT

The authors thank Dr. Anil Kumar Sharma, Professor and Head, Department of Biotechnology, Maharishi Markandeshwar (Deemed to be University), Mullana, Ambala for his kind support to carry out this study.

## CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

## ABBREVIATIONS

**CFU:** Colony Forming Unit; **COPD:** Chronic Obstructive Pulmonary Disease; **CVD:** Cardio Vascular Disease; **DAEC:** Diffusely Adherent *E. coli*; **DNA:** Deoxyribo-Nucleic Acid; **EAEC:** Entero-Aggregative *E. coli*; **EHEC:** Entero-Haemorrhagic *E. coli*; **EPEC:** Entero-Pathogenic *E. coli*; **ETEC:** Entero-Toxigenic *E. coli*; **GNB:** Gram Negative Bacteria; **GPB:** Gram Positive Bacteria; **MALDI-TOF MS:** Matrix-Assisted Laser Desorption Ionization-Time of Flight Mass Spectrometry; **QMRA:** Quantitative Microbial Risk Assessment; **RTE:** Ready To Eat; **SPC:** Standard Plate

Count; **TVC:** Total Viable Count; **WHO:** World Health Organization.

## SUMMARY

To summarize the present study, an array of microbial flora associated with vegetable salads was discussed in this study. Risk factors and levels of microbiological quality satisfaction are also discussed. Different phenotypic and molecular methods were discussed for correct identification of contaminating microbes. Good hand hygiene is highly recommended along with proper washing of vegetable salads before consumption.

## REFERENCES

1. Beuchat LR. Ecological factors influencing survival and growth of human pathogens on raw fruits and vegetables. *Microbes Infect.* 2002;4(4):413-23. doi: 10.1016/s1286-4579(02)01555-1, PMID 11932192.
2. Li J, Wang Z, Karim MR, Zhang L. Detection of human intestinal protozoan parasites in vegetables and fruits: A review. *Parasit Vectors.* 2020;13(1):380. doi: 10.1186/s13071-020-04255-3, PMID 32727529.
3. Van Dijken GD, Uijtewaal PH, Logtenberg SJJ, Sankatsing SUC. [Scombroid food poisoning among hospital personnel]. *Ned Tijdschr Geneesk.* 2020;164. PMID 32613785.
4. Ibrahim JN, Eghnatio E, El Roz A, Fardoun T, Ghssein G. Prevalence, antimicrobial resistance and risk factors for campylobacteriosis in Lebanon. *J Infect Dev Ctries.* 2019;13(1):11-20. doi: 10.3855/jidc.10729, PMID 32032018.
5. Thike TZ, Saw YM, Lin H, Chit K, Tun AB, Htet H, *et al.* Association between body mass index and ready-to-eat food consumption among sedentary staff in Nay Pyi Taw union territory, Myanmar. *BMC Public Health.* 2020;20(1):206. doi: 10.1186/s12889-020-8308-6, PMID 32041555.
6. Chonpracha P, Ardoin R, Gao Y, Waimaleongora-Ek P, Tuuri G, Prinyawiwatkul W. Effects of intrinsic and extrinsic visual cues on consumer emotion and purchase intent: A case of ready-to-eat salad. *Foods.* 2020;9(4). doi: 10.3390/foods9040396, PMID 32244291.
7. Wallace TC, Bailey RL, Blumberg JB, Burton-Freeman B, Chen CO, Crowe-White KM, *et al.* Fruits, vegetables, and health: A comprehensive narrative, umbrella review of the science and recommendations for enhanced public policy to improve intake. *Crit Rev Food Sci Nutr.* 2020;60(13):2174-211. doi: 10.1080/10408398.2019.1632258, PMID 31267783.
8. Springmann M, Spajic L, Clark MA, Poore J, Herforth A, Webb P, *et al.* The healthiness and sustainability of national and global food based dietary guidelines: Modelling study. *BMJ.* 2020;370:m2322. doi: 10.1136/bmj.m2322, PMID 32669369.
9. Jayawardena R, Jeyakumar DT, Gamage M, Sooriyaarachchi P, Hills AP. Fruit and vegetable consumption among South Asians: A systematic review and meta-analysis. *Diabetes Metab Syndr.* 2020;14(6):1791-800. doi: 10.1016/j.dsx.2020.09.004, PMID 32947110.
10. Zhou T, Wang M, Ma H, Li X, Heianza Y, Qi L. Dietary fiber, genetic variations of gut microbiota-derived short-chain fatty acids, and bone health in UK Biobank. *J Clin Endocrinol Metab.* 2021;106(1):201-10. doi: 10.1210/clinem/dgaa740, PMID 33051670.
11. Xylia P, Botsaris G, Chrysargyris A, Skandamis P, Tzortzakis N. Variation of microbial load and biochemical activity of ready-to-eat salads in Cyprus as affected by vegetable type, season, and producer. *Food Microbiol.* 2019;83:200-10. doi: 10.1016/j.fm.2019.05.013, PMID 31202414.
12. Minich DM, Brown BI. A review of dietary (phyto)nutrients for glutathione support. *Nutrients.* 2019;11(9). doi: 10.3390/nu11092073, PMID 31484368.
13. Dayib M, Larson J, Slavin J. Dietary fibers reduce obesity-related disorders: Mechanisms of action. *Curr Opin Clin Nutr Metab Care.* 2020;23(6):445-50. doi: 10.1097/MCO.0000000000000696, PMID 32925180.

14. Anderson JW, Baird P, Davis RH, Jr., Ferreri S, Knudtson M, Koraym A, *et al.* Health benefits of dietary fiber. *Nutr Rev.* 2009;67(4):188-205. doi: 10.1111/j.1753-4887.2009.00189.x, PMID 19335713.
15. Yang JJ, Yu D, Xiang YB, Blot W, White E, Robien K, *et al.* Association of dietary fiber and yogurt consumption with lung cancer risk: A pooled analysis. *JAMA Oncol.* 2020;6(2):e194107. doi: 10.1001/jamaoncol.2019.4107, PMID 31647500.
16. Farvid MS, Spence ND, Holmes MD, Barnett JB. Fiber consumption and breast cancer incidence: A systematic review and meta-analysis of prospective studies. *Cancer.* 2020;126(13):3061-75. doi: 10.1002/cncr.32816, PMID 32249416.
17. Srour B, Fezeu LK, Kesse-Guyot E, Allès B, Méjean C, Andrianasolo RM, *et al.* Ultra-processed food intake and risk of cardiovascular disease: prospective cohort study (NutriNet-Sante). *BMJ.* 2019;365:l1451. doi: 10.1136/bmj.l1451, PMID 31142457.
18. Van Duyn MA, Pivonka E. Overview of the health benefits of fruit and vegetable consumption for the dietetics professional: Selected literature. *J Am Diet Assoc.* 2000;100(12):1511-21. doi: 10.1016/S0002-8223(00)00420-X, PMID 11138444.
19. Vieira AR, Abar L, Vingeliene S, Chan DS, Aune D, Navarro-Rosenblatt D, *et al.* Fruits, vegetables and lung cancer risk: A systematic review and meta-analysis. *Ann Oncol.* 2016;27(1):81-96. doi: 10.1093/annonc/mdv381, PMID 26371287.
20. Ávila-Gálvez MÁ, Giménez-Bastida JA, Espín JC, González-Sarrias A. Dietary phenolics against breast cancer. A critical evidence-based review and future perspectives. *Int J Mol Sci.* 2020;21(16). doi: 10.3390/ijms21165718, PMID 32784973.
21. Yuan K, Zhu Q, Lu Q, Jiang H, Zhu M, Li X, *et al.* Quercetin alleviates rheumatoid arthritis by inhibiting neutrophil inflammatory activities. *J Nutr Biochem.* 2020;84:108454. doi: 10.1016/j.jnutbio.2020.108454.
22. Oishi Y, Imamura T, Shimomura T, Suzuki K. Vegetable freshness perception in dementia with Lewy bodies and Alzheimer's disease. *Dement Geriatr Cogn Dis Extra.* 2020;10(2):74-85. doi: 10.1159/000508282, PMID 33082771.
23. Park HA, Hayden MM, Bannerman S, Jansen J, Crowe-White KM. Anti-apoptotic effects of carotenoids in neurodegeneration. *Molecules.* 2020;25(15). doi: 10.3390/molecules25153453, PMID 32751250.
24. Block G, Patterson B, Subar A. Fruit, Vegetables, and cancer prevention: a review of the epidemiological evidence. *Nutr Cancer.* 1992;18(1):1-29. doi: 10.1080/01635589209514201, PMID 1408943.
25. Klinder A, Shen Q, Heppel S, Lovegrove JA, Rowland I, Tuohy KM. Impact of increasing fruit and vegetables and flavonoid intake on the human gut microbiota. *Food Funct.* 2016;7(4):1788-96. doi: 10.1039/c5fo01096a, PMID 26757793.
26. Anderson JW, Smith BM, Gustafson NJ. Health benefits and practical aspects of high-fiber diets. *Am J Clin Nutr.* 1994;59(5);Suppl:1242S-7S. doi: 10.1093/ajcn/59.5.1242S, PMID 8172129.
27. Fulgoni VL, 3<sup>rd</sup>. Wallace TC, Stylianou KS, Jolliet O. Calculating intake of dietary risk components used in the global burden of disease studies from the what we eat in America/national health and nutrition examination surveys. *Nutrients.* 2018;10(10). doi: 10.3390/nu10101441, PMID 30301145.
28. Ramos-Ramírez LDC, Romero-Bañuelos CA, Jiménez-Ruiz EI, Palomino-Hermosillo YA, Saldaña-Ahuactzi Z, Martínez-Laguna Y, *et al.* Coliform bacteria in San Pedro Lake, western Mexico. *Water Environ Res.* 2021;93(3):384-92. doi: 10.1002/wer.1423, PMID 32757433.
29. Herrero-Fresno A, Olsen JE. Effect of ampicillin, cephalixin, ceftiofur and tetracycline treatment on selection of resistant coliforms in a swine faecal microcosmos. *J Appl Microbiol.* 2020;129(5):1238-47. doi: 10.1111/jam.14721, PMID 32430970.
30. McInnes RS, Uz-Zaman MH, Alam IT, Ho SFS, Moran RA, Clemens JD, *et al.* Metagenome-wide analysis of rural and urban surface waters and sediments in Bangladesh identifies human waste as a driver of antibiotic resistance. *mSystems.* 2021;6(4):e0013721. doi: 10.1128/mSystems.00137-21, PMID 34254820.
31. Cho GS, Stein M, Fiedler G, Igbinosa EO, Koll LP, Brinks E, *et al.* Polyphasic study of antibiotic-resistant enterobacteria isolated from fresh produce in Germany and description of *Enterobacter vonholyi* sp. nov. isolated from marjoram and *Enterobacter dykesii* sp. nov. isolated from mung bean sprout. *Syst Appl Microbiol.* 2021;44(1):126174. doi: 10.1016/j.syapm.2020.126174.
32. Salazar-Llorente E, Morales M, Sornoza I, Mariduena-Zavala MG, Gu G, Nou X, *et al.* Microbiological quality of high-demand food from three major cities in Ecuador. *J Food Prot.* 2021;84(1):128-38. doi: 10.4315/JFP-20-271, PMID 33411929.
33. Atherholt TB, Procopio NA, Goodrow SM. Seasonality of coliform bacteria detection rates in New Jersey domestic wells. *Ground Water.* 2017;55(3):346-61. doi: 10.1111/gwat.12482, PMID 27775834.
34. Nowicki S, deLaurent ZR, De Villiers EP, Githinji G, Charles KJ. The utility of *Escherichia coli* as a contamination indicator for rural drinking water: Evidence from whole genome sequencing. *PLOS ONE.* 2021;16(1):e0245910. doi: 10.1371/journal.pone.0245910, PMID 33481909.
35. Wang Y, Xu C, Zhang R, Chen Y, Shen Y, Hu F, *et al.* Changes in colistin resistance and *mcr-1* abundance in *Escherichia coli* of animal and human origins following the ban of colistin-positive additives in China: an epidemiological comparative study. *Lancet Infect Dis.* 2020;20(10):1161-71. doi: 10.1016/S1473-3099(20)30149-3, PMID 32505232.
36. Wight J, Varin MP, Robertson GJ, Huot Y, Lang AS. Microbiology in the field: construction and validation of a portable incubator for real-time quantification of coliforms and other bacteria. *Front Public Health.* 2020;8:607997. doi: 10.3389/fpubh.2020.607997.
37. Al Nahhas S, Aboualchamat G. Investigation of parasitic contamination of salad vegetables sold by street vendors in city markets in Damascus, Syria. *Food Waterborne Parasitol.* 2020;21:e00090. doi: 10.1016/j.fawpar.2020.e00090, PMID 33241130.
38. Lalonde LF, Xie V, Oakley JR, Lobanov VA. Optimization and validation of a loop-mediated isothermal amplification (LAMP) assay for detection of *Giardia duodenalis* in leafy greens. *Food Waterborne Parasitol.* 2021;23:e00123. doi: 10.1016/j.fawpar.2021.e00123, PMID 34169158.
39. Pinto-Ferreira F, Reis JB, Paschoal ATP, Balbino LS, Bertão-Santos A, Lucas JI, *et al.* Molecular diagnosis of the curly lettuce parasitic contamination from hydroponic cultivation from supermarkets. *Rev Bras Parasitol Vet.* 2020;29(4):e015820. doi: 10.1590/S1984-296120200095, PMID 33237193.
40. Hajipour N, Soltani M, Ketzis J, Hassanzadeh P. Zoonotic parasitic organisms on vegetables: Impact of production system characteristics on presence, prevalence on vegetables in northwestern Iran and washing methods for removal. *Food Microbiol.* 2021;95:103704. doi: 10.1016/j.fm.2020.103704.
41. Akoachere JTK, Tatsinkou BF, Nkengfack JM. Bacterial and parasitic contaminants of salad vegetables sold in markets in Fako Division, Cameroon and evaluation of hygiene and handling practices of vendors. *BMC Res Notes.* 2018;11(1):100. doi: 10.1186/s13104-018-3175-2, PMID 29409524.
42. Stromberg ZR, Johnson JR, Fairbrother JM, Kilbourne J, Van Goor A, Curtiss RR, *et al.* Evaluation of *Escherichia coli* isolates from healthy chickens to determine their potential risk to poultry and human health. *PLOS ONE.* 2017;12(7):e0180599. doi: 10.1371/journal.pone.0180599, PMID 28671990.
43. Hassan R, Seelman S, Peralta V, Booth H, Tewell M, Melius B, *et al.* A multistate outbreak of *E. coli* O157:H7 infections linked to soy nut butter. *Pediatrics.* 2019;144(4). doi: 10.1542/peds.2018-3978, PMID 31519792.
44. Gomes TA, Elias WP, Scaletsky IC, Guth BE, Rodrigues JF, Piazza RM, *et al.* Diarrheagenic *Escherichia coli*. *Braz J Microbiol.* 2016;47;Suppl 1:3-30. doi: 10.1016/j.bjm.2016.10.015, PMID 27866935.
45. Manges AR, Geum HM, Guo A, Edens TJ, Fibke CD, Pitout JDD. Global extraintestinal Pathogenic *Escherichia coli* (ExPEC) lineages. *Clin Microbiol Rev.* 2019;32(3). doi: 10.1128/CMR.00135-18, PMID 31189557.
46. Weiner LM, Webb AK, Limbago B, Dudeck MA, Patel J, Kallen AJ, *et al.* Antimicrobial-resistant pathogens associated with healthcare-associated infections: summary of data reported to the national healthcare safety network at the Centers for Disease Control and Prevention, 2011-2014. *Infect Control Hosp Epidemiol.* 2016;37(11):1288-301. doi: 10.1017/ice.2016.174, PMID 27573805.
47. Waturangi DE, Hudiono F, Aliwarga E. Prevalence of pathogenic *Escherichia coli* from salad vegetable and fruits sold in Jakarta. *BMC Res Notes.* 2019;12(1):247. doi: 10.1186/s13104-019-4284-2, PMID 31046825.
48. Schwidder M, Heinisch L, Schmidt H. Genetics, Toxicity, and Distribution of enterohemorrhagic *Escherichia coli* Hemolysin. *Toxins (Basel).* 2019;11(9). doi: 10.3390/toxins11090502, PMID 31470552.
49. Van den Beld MJC, Reubsat FAG, Pijnacker R, Harpal A, Kuiling S, Heerkens EM, *et al.* A multifactorial approach for surveillance of *Shigella* spp.

- and entero-invasive *Escherichia coli* is important for detecting (inter)national clusters. *Front Microbiol.* 2020;11:564103. doi: 10.3389/fmicb.2020.564103.
50. Meza-Segura M, Zaidi MB, Vera-Ponce de León A, Moran-García N, Martínez-Romero E, Nataro JP, *et al.* New insights into DAEC and EAEC pathogenesis and phylogeny. *Front Cell Infect Microbiol.* 2020;10:572951. doi: 10.3389/fcimb.2020.572951.
  51. Chau ML, Aung KT, Hapuarachchi HC, Lee PS, Lim PY, Kang JS, *et al.* Microbial survey of ready-to-eat salad ingredients sold at retail reveals the occurrence and the persistence of *Listeria monocytogenes* Sequence Types 2 and 87 in pre-packed smoked salmon. *BMC Microbiol.* 2017;17(1):46. doi: 10.1186/s12866-017-0956-z, PMID 28245788.
  52. Abaza A. Bacteriological assessment of some vegetables and ready-to-eat salads in Alexandria, Egypt. *J Egypt Public Health Assoc.* 2017;92(3):177-87. doi: 10.21608/EPX.2018.16151, PMID 30341996.
  53. Arienzo A, Murgia L, Fraudentali I, Gallo V, Angelini R, Antonini G. Microbiological quality of ready-to-eat leafy green salads during shelf-life and home-refrigeration. *Foods.* 2020;9(10). doi: 10.3390/foods9101421, PMID 33049952.
  54. Elpers L, Kretschmar J, Nuccio SP, Bäumlér AJ, Hensel M. Factors required for adhesion of *Salmonella enterica* serovar Typhimurium to corn salad (*Valerianella locusta*). *Appl Environ Microbiol.* 2020;86(8). doi: 10.1128/AEM.02757-19, PMID 32033951.
  55. Kadariya J, Smith TC, Thapaliya D. *Staphylococcus aureus* and staphylococcal food-borne disease: an ongoing challenge in public health. *BioMed Res Int.* 2014;2014:827965. doi: 10.1155/2014/827965.
  56. Aycicek H, Oguz U, Karci K. Determination of total aerobic and indicator bacteria on some raw eaten vegetables from wholesalers in Ankara, Turkey. *Int J Hyg Environ Health.* 2006;209(2):197-201. doi: 10.1016/j.ijheh.2005.07.006, PMID 16503303.
  57. Younus MI, Sabuj AAM, Haque ZF, Sayem SM, Majumder S, Parvin MS, *et al.* Microbial risk assessment of ready-to-eat mixed vegetable salads from different restaurants of Bangladesh Agricultural University campus. *J Adv Vet Anim Res.* 2020;7(1):34-41. doi: 10.5455/javar.2020.g390, PMID 32219107.
  58. Duffy G, Cummins E, Nally P, O'Brien S, Butler F. A review of quantitative microbial risk assessment in the management of *Escherichia coli* O157:H7 on beef. *Meat Sci.* 2006;74(1):76-88. doi: 10.1016/j.meatsci.2006.04.011, PMID 22062718.
  59. Cassin MH, Lammerding AM, Todd EC, Ross W, McColl RS. Quantitative risk assessment for *Escherichia coli* O157:H7 in ground beef hamburgers. *Int J Food Microbiol.* 1998;41(1):21-44. doi: 10.1016/s0168-1605(98)00028-2, PMID 9631335.
  60. Stephen JF. The microbiological risk assessment of food. UK: Department of Life Sciences, Nottingham Trent University. Wiley-Blackwell; 2002.
  61. Moyes RB, Reynolds J, Breakwell DP. Differential staining of bacteria: Gram stain. *Curr Protoc Microbiol.* 2009;Appendix(3):Appendix 3C. doi: 10.1002/9780471729259.mca03cs15, PMID 19885931; Appendix 3: Appendix 3C.
  62. Tsuchida S, Umemura H, Nakayama T. Current status of matrix-assisted laser desorption/ionization-time-of-flight mass spectrometry (MALDI-TOF MS) in clinical diagnostic microbiology. *Molecules.* 2020;25(20). doi: 10.3390/molecules25204775, PMID 33080897.
  63. Jang KS, Kim YH. Rapid and robust MALDI-TOF MS techniques for microbial identification: a brief overview of their diverse applications. *J Microbiol.* 2018;56(4):209-16. doi: 10.1007/s12275-018-7457-0, PMID 29492868.
  64. El-Nemr IM, Mushaha M, Sundararaju S, Fontejon C, Suleiman M, Tang P, *et al.* Application of MALDI biotyper system for rapid identification of bacteria isolated from a fresh produce market. *Curr Microbiol.* 2019;76(3):290-6. doi: 10.1007/s00284-018-01624-1, PMID 30603962.

**Cite this article:** Kumar S, Yadav M, Devi A, Uniyal M, Kumar V, Sehrawat N, Singh R. Assessment of Pathogenic Microorganisms Associated with Vegetable Salads. *Asian J Biol Life Sci.* 2022;11(1):1-7.